

# Convergent evolution of floral shape tied to pollinator shifts in lochrominae (Solanaceae)\*

Stacey D. Smith<sup>1,2</sup> and Ricardo Kriebel<sup>3</sup>

<sup>1</sup>Department of Ecology and Evolutionary Biology, University of Colorado, Boulder Colorado 80305 <sup>2</sup>E-mail: Stacey.D.Smith@colorado.edu <sup>3</sup>Department of Botany, University of Wisconsin – Madison, Madison, Wisconsin 53706

Received August 2, 2017 Accepted December 8, 2017

Flower form is one of many floral features thought to be shaped by pollinator-mediated selection. Although the drivers of variation in flower shape have often been examined in microevolutionary studies, relatively few have tested the relationship between shape evolution and shifts in pollination system across clades. In the present study, we use morphometric approaches to quantify shape variation across the Andean clade lochrominae and estimate the relationship between changes in shape and shifts in pollination system using phylogenetic comparative methods. We infer multiple shifts from an ancestral state of narrow, tubular flowers toward open, bowl-shaped, or campanulate flowers as well as one reversal to the tubular form. These transitions in flower shape are significantly correlated with changes in pollination system. Specifically, tubular forms tend to be hummingbird-pollinated and the open forms tend to be insect-pollinated, a pattern consistent with experimental work as well as classical floral syndromes. Nonetheless, our study provides one of the few empirical demonstrations of the relationship between flower shape and pollination system at a macroevolutionary scale.

**KEY WORDS:** Floral syndromes, morphometrics, PGLS, pollinator importance.

Although the variation in form, shape, and color of flowers has often been attributed to pollinator-mediated selection, the nature of this process and its importance in driving divergence remain contentious. Various authors have argued for the pollinator-shift model (the Grant-Stebbins model; Johnson 2006), wherein suites or "syndromes" of floral traits are shaped by "functional" groups of pollinators, and floral diversity arises through shifts among these functional groups (Grant 1949; Stebbins 1970; Fenster et al. 2004). While the pollinator-shift model appears to fit well with the floral variation and pollination systems of many classic radiations (e.g., Wilson et al. 2004; Whittall and Hodges 2007; Tripp and Manos 2008), it remains unclear how well shifts among major groups of pollinators explain diversity more broadly (Van der Niet et al. 2014). For example, many florally diverse groups of plants have radiated while retaining an essentially unchanged

\*This article corresponds to Dodsworth, S., A. Orejuela, O. A. Pérez-Escobar, T. Särkinen, and S. Knapp. 2018. Digest: Shape-shifting in Solanaceae flowers: The influence of pollinators. Evolution. https://doi.org/ 10.1111/evo.13437. but specialized pollination ecology (De Luca and Vallejo-Marin 2013; Davis et al. 2014). Pollinators may still play a role in floral divergence in these systems, for example through competitive interactions (Armbruster et al. 1994; Muchhala and Potts 2007; Eaton et al. 2012). Equally, many factors beyond pollinators, such as herbivores and climatic features, are known to influence floral evolution (Armbruster 2002; Strauss and Whittall 2006).

Testing the relationship between pollinator shifts and floral trait evolution on a macroevolutionary scale is largely hindered by the lack of comparative quantitative data. Although broad-scale studies often treat pollination system and floral traits as discrete characters (e.g., Ollerton et al. 2009; Rosas-Guerrero et al. 2014), both vary continuously in natural systems (Waser et al. 1996; Barrett and Hodgins 2006; Conner 2006) and are thus best treated as continuous traits. Nonetheless, capturing floral diversity and plant–pollinator interactions as continuous variables amenable to analysis can be challenging. In terms of quantifying the contribution of a pollinator to a plant's reproduction, metrics such as pollinator efficiency and pollinator importance have been proposed

to incorporate aspects of both visit frequency ("quantity") and the amount of pollen deposited ("quality") (reviewed in Ne'eman et al. 2010). Easily measured floral traits, such as aspects of length and width, have been commonly studied in the context of pollinator shifts across clades (Wilson et al. 2004; Whittall and Hodges 2007) while complex traits such as patterning, reward, and shape have received less attention (but see Armbruster 1990; Perez et al. 2006; Gomez et al. 2008). Nonetheless, recent advances in geometric morphometrics (reviewed in Klingenberg 2010) have reinvigorated the study of floral shape and the role of pollinators in its evolution (van der Niet et al. 2010; Fernandez-Mazuecos et al. 2013; Gomez et al. 2014).

While macroevolutionary tests of pollinator-mediated floral shape evolution remain uncommon, studies of within-species variation provide insight into the mechanisms of selection. For example, pollinators may exhibit differential visitation rates with respect to flower shape variation, resulting in directional selection (e.g., Galen and Cuba 2001; Gomez et al. 2006). This apparent pollinator preference can arise from a variety of factors, including differences in the rate of approach, handling time, or correlated traits, such as nectar reward (Cresswell and Galen 1991; Sutherland and Vickery 1996; Fenster et al. 2006; Temeles et al. 2009). Shape variation may also influence the interaction between the pollinator and the floral reproductive organs, leading to associated variation in pollen deposition or removal. This link between flower shape and pollinator efficiency has well-documented consequences for both male and female components of fitness (e.g., Campbell et al. 1996; Kulbaba and Worley 2013; La Rosa and Conner 2017). Importantly in the context of the pollinator-shift model, different pollinators often exert different selective pressures on flower shape (Campbell 2004; Muchhala 2007; Gomez et al. 2008), suggesting that shifts among pollinators, whether within a single species or across a clade, will lead to corresponding shifts in multiple aspects of shape.

Here, we quantify the diversity of corolla shapes in Iochrominae and assess the relationship between shape variation and pollination system across this clade. Iochrominae comprises roughly 35 Andean species in the tomato family (Solanaceae). It has been traditionally divided among seven small genera and is known for its tremendous diversity of flower shapes, sizes, and colors (Smith and Baum 2006). Previous studies have shown that a wide array of insects and hummingbirds can act as effective pollinators (Smith et al. 2008b). Most of the species of Iochroma, the largest genus, are principally pollinated by hummingbirds, while some are pollinated by a mix of hummingbirds and insects and those in closely related genera are often principally insect pollinated (Taura and Laroca 2004; Verçoza et al. 2012). These differences in pollination system are closely associated with reward and display, as species with large displays and high-reward flowers are more likely to be pollinated by hummingbirds (Smith et al. 2008a).

Combining this existing body of information about pollination ecology of Iochroma and its relatives with new floral morphometrics and an expanded phylogeny, we test whether pollinator shifts are associated with floral shape evolution. Iochrominae are radially symmetric (or at most weakly bilaterally symmetric) along the dorsal-ventral axis (Knapp 2010), but they vary greatly along the proximal-distal axis, with shapes ranging from widely campanulate to narrowly tubular (Smith and Baum 2007). We hypothesize that this shape variation is tied to transitions between insect, bird, and mixed pollination systems, with narrow flowers being favored in bird-pollinated taxa and open flowers in insectpollinated taxa. In the present study, we apply geometric morphometrics to quantify corolla shape variation and conduct statistical comparative analyses to estimate its relationship to changes in pollination system. In addition, we use the phylogeny to trace the evolution of shape and determine the extent of shape convergence across the clade.

# Materials and Methods PHYLOGENY

To reconstruct the evolution of flower shape, we updated the existing calibrated phylogeny of Iochrominae (Smith and Goldberg 2015) to include a recently discovered species of Saracha (S. andina Rob. Fernandez, I. Revilla and E. Pariente). We added S. andina sequences from Fernandez-Hilario and Smith (2017) to the existing three-gene alignment, which includes the internal transcribed spacer (ITS), LFY intron II, and exons 2 through 9 of waxy. In total, we sampled 36 ingroup species (Fig. 1A) and included six additional Solanaceae as outgroups, following Fernandez-Hilario and Smith (2017). We conducted likelihood ratio tests in PAUP 4.0b10 (Swofford 2002) to compare the fit of five models (JC, K2P, HKY, GTR, and GTR+ $\Gamma$ ) and chose the best model (GTR+  $\Gamma$ ) for phylogenetic inference. To obtain ultrametric trees suitable for comparative methods, we estimated the phylogeny using relaxed-clock methods in BEAST v. 2.1.2 (Bouckaert et al. 2014). Following Smith and Goldberg (2015), we used a relaxed-clock log-normal model for rate variation across branches, a birth-death model for trees, a set of three secondary calibrations (one for the root node, one for the Iochrominae-Physalinae clade, and one for Iochrominae) from Särkinen et al. (2013). We completed two runs of 5 million generations, and after evaluating convergence and effective sample size, we removed the first 25% of trees from as burn-in. We computed the maximum clade-credibility (MCC) tree using the TreeAnnotator program in BEAST. We also subsampled 100 trees from the posterior distribution for downstream analyses. Outgroups were pruned from the tree before analyses. The final alignment, MCC tree (Fig. S1) and 100-tree sample are deposited in Dryad (https://doi.org/10.5061/dryad.5jn7b).



**Figure 1.** Iochrominae phylogeny and floral morphospace. (A) MCC tree from BEAST analysis of the three-gene dataset for lochrominae. Abbreviations for genera of lochrominae are I. = *lochroma*, *A*. = *Acnistus*, *V*. = *Vassobia*, *S*. = *Saracha*, *E*. = *Eriolarynx*, *D*. = *Dunalia*. Outgroups (*Tubocapsicum anomalum*, *Physalis peruviana*, *Witheringia solanacea*, *Leucophysalis grandiflora*, *Cuatresia harlingiana*, and *Larnax sachapapa*) were pruned from the tree. Major clades ("A," "CLF," "DESV," "U") are labeled following Smith and Baum (2006). (B) Shape variation along PC axes. For each of the first two PCs, the mean is shown along with the shapes corresponding to ±1 standard deviation (SD). (C) Empirical morphospace with outlines for each of the 36 species. Colored dots correspond to clade membership shown in (A).

## **MORPHOMETRIC ANALYSIS**

Corolla shape was characterized using images of fresh flowers taken in the field. For each species, a single mature flower from the individual sampled for the phylogeny was photographed in lateral view, providing a two-dimensional representation of the shape along the proximal-distal axis. These images were converted into silhouettes in Adobe Photoshop (San Jose, CA), and the silhouettes were transformed into outlines using the R package Momocs (Bonhomme et al. 2014). Each outline was converted into a list of 200 x, y coordinates describing the polygon and then centered, also using Momocs. Variation in shape was quantified using elliptical Fourier analysis (EFA), which decomposed the outlines into 32 harmonics, each with four coefficients. To avoid twisting of the outlines during the EFA, four landmarks were placed on each outline, one on each side of each tube, and one above and one below in the middle area of each tube. These landmarks were then used to align the outlines

with a Full Generalized Procrustes alignment in Momocs. The 128 coefficients resulting from the EFA were summarized using a principal component analysis (PCA). Species were plotted along the PC axes to visualize their distribution in morphospace. As an indication of phylogenetic structure in these data, we computed Blomberg's K for the PC variables (Blomberg et al. 2003) and used 1000 randomizations of the data across the MCC tree to test if the value was significantly different from K = 0 (no phylogenetic signal). Scripts and shape outlines used for all analyses are deposited in Dryad (https://doi.org/10.5061/dryad.5jn7b).

# ASSOCIATION BETWEEN SHAPE VARIATION AND POLLINATION SYSTEM

We used phylogenetic generalized least squares (PGLS) to test the relationship between shape variation and pollination system. Pollination system was characterized using pollinator importance, a metric that incorporates both visitation rates and effectiveness in terms of pollen deposition (Waser and Price 1983; Mayfield et al. 2001). Estimates of pollinator importance for both insects and hummingbird visitors are available for 15 species of Iochrominae from previous studies (Smith et al. 2008b). We also included data from Taura and Laroca (2004), who studied the pollination of *Vassobia breviflora* during two flowering seasons. Given the small size of this dataset (16 species), we focused our analyses on relative hummingbird importance, which ranges from 0 to 1, with 1 being 100% bird-pollinated. Since insects constitute the non-hummingbird pollinators, relative hummingbird importance in this case can be considered 1 minus relative insect importance. *V. breviflora* was scored as 0 given that it is exclusively beepollinated (Taura and Laroca 2004), and data for the remaining species was taken from Smith et al. (2008b).

We regressed shape PCs from the morphometric analysis against values for pollinator importance with phylogeneticgeneralized least squares (PGLS, Martins and Hansen 1997) using the gls functions in the nlme package in R (Pinheiro et al. 2017). We assumed a correlation structure based on the Ornstein-Uhlenbeck (OU) process, which models stabilizing selection around an optimum (Hansen 1997; Butler and King 2004). The strength of selection is determined by the  $\alpha$  parameter, which we estimated during model fitting. When  $\alpha$  is zero, the model is equivalent to Brownian motion and as  $\alpha$  increases toward infinity, the model converges on a non-phylogenetic "TIPS" model. Given that multiple evolutionary processes can result in the similar patterns of phylogenetic structure in the data (Revell et al. 2008), we consider this approach primarily as a way to adjust for the level of statistical dependence among the trait values due to the phylogeny. The PGLS analyses were repeated for the first two shape PCs using the MCC tree and also the sample of 100 trees from the posterior distribution. The  $\alpha$  parameter was optimized for each tree during PGLS model fitting. Distributions for each parameter across the 100 trees were summarized by computing the median and the range.

Given the relatively small number of taxa in this data, we implemented several approaches to examine the robustness of our results. First, we repeated the PGLS analyses excluding *Vassobia breviflora*, the species whose pollination was not studied directly by the first author. It appears as an outlier in the shape analysis (see Results, Morphospace), having a much wider mouth than the other 15 species for which pollinator data is available. Second, we conducted a phylogenetic ANOVA analysis, treating pollination system as a categorical variable. We included six additional taxa for which there were field observations of pollinators (Table S1), but no quantitative study. Pollination systems were categorized as principally hummingbird-pollinated ( $\geq$ 75% relative hummingbird importance for species with quantitative data or only hummingbirds observed for those without), principally insect-pollinated ( $\geq$ 75% relative insect

importance or only insects observed), or mixed (intermediate values of pollinator importance or both insect and hummingbird visits observed). Third, these PGLS and phylogenetic ANOVA analyses were repeated using multiple structures to model the degree of phylogenetic correlation in the residuals (the Brownian motion (BM) model, the OU model, and the TIPS model).

#### PATTERNS OF CONVERGENCE

Given the relationship between the first corolla shape PC and pollinator importance detected in the PGLS analysis, we applied model-based methods to examine the extent of convergence in this aspect of floral shape. We first constructed a traitgram for PC1 using the MCC tree with the phenogram function in phytools (Revell 2012). We then used the llou R package (Khabbazian et al. 2016) to estimate the number of distinct optima (or selective regimes) across the phylogeny under an Ornstein-Uhlenbeck process using a LASSO (Least Absolute Shrinkage and Selection Operator) procedure (Tibshirani 1996). One of the advantages of 11ou is that it does not require the a priori designation of where regime shifts may have occurred. Convergence is inferred with 11ou as independent shifts to the same regime. We chose to use the phylogenetic Bayesian information criterion (pBIC) for model selection as it has been shown to minimize the inference of false shifts (Khabbazian et al. 2016). We repeated these analyses across the sample of 100 trees from the posterior distribution to assess the effect of phylogenetic uncertainty on the results.

## Results morphospace

The PCA analysis of harmonic coefficients from the elliptic Fourier analysis captured 91.2% of the variation in the first two principal components. Principal component 1 accounts for 74.7% of the variation and describes the narrowness of the tube and to some degree, the extent of constriction of the tube toward the middle (Fig. 1B). The second principal component, comprising 16.4% of the variation, captures the amount of restriction at the mouth, given that some species have strongly flared corollas and other are relatively straight. Tubular shapes with slightly flared mouths appear to be the most common forms and are characteristic of the largest genera, *Iochroma* and *Dunalia*.

Although corolla shapes were clustered to some extent by relatedness, the phylogenetic distribution suggests multiple shifts among the various forms. For example, bowl-shaped flowers are only found in the two sister species of *Vassobia* (Fig. 1C, upper right), but funnel-shaped flowers are found in several clades and traditional genera (*Acnistus, Saracha*, and *Iochroma*). Similarly, tubular forms appear in all of the major clades. Accordingly, species with similar flower shapes are often distantly related.

Model	Tree	Coefficient	SE	α	Р	σ
PC1~ Poll	MCC	0.29	0.08	0.18	0.004	0.16
	100 trees	0.28	0.08	1.31	0.004	0.14
		(0.27, 0.32)	(0.08, 0.09)	(0.00, 7.21)	(0.001, 0.006)	(0.12, 584)
$PC2 \sim Poll$	MCC	-0.01	0.05	0.00	0.86	
	100 trees	-0.02	0.05	0.08	0.74	330
		(-0.03, 0.00)	(0.04, 0.05)	(0.00, 1.34)	(0.46, 0.98)	(0.09, 2255)

Table 1. Parameter estimates from PGLS analyses.

"Poll" denotes relative hummingbird importance. For PGLS analyses across the 100 tree samples, median parameter values are presented with 95% intervals in parentheses.

Blomberg's *K* suggests little phylogenetic signal (K = 0.30, P = 0.63) in the first shape PC. The second PC does show significant phylogenetic signal (K = 0.78, P = 0.001), but this component accounts for a small portion of the variation in shape compared to PC1.

## ASSOCIATION BETWEEN SHAPE VARIATION AND POLLINATION SYSTEM

PGLS analyses indicated a strong relationship between pollination system and the first shape PC but not the second. Species with constricted, tubular corollas (higher PC1 scores) tend to be pollinated by hummingbirds while those with open corollas tend to be pollinated by a mix of hummingbirds and insects, or insects exclusively (Table 1; Fig. 2). One species, *I. lehmannii* appears as an outlier with more bird pollination that would be expected based on its shape, suggesting that additional factors are likely at play in this taxon. The significant relationship between PC1 and relative hummingbird importance was recovered in all of the replicated PGLS analyses across the 100 Bayesian trees (P = 0.001-0.006, Table 1). By contrast, PC2 was not associated with pollination system in any analysis (P = 0.46-0.98, Table 1).

Our sensitivity analyses suggested that these associations are robust, despite the small sample size in the original dataset (16 species). The relationship between shape PC1 and pollinator importance was recovered in PGLS analyses with all three models (BM, OU, TIPS) after removing the *V. breviflora* outlier, albeit with reduced significance levels (P = 0.01-0.03, Table S2). The phylogenetic ANOVA with pollination system coded as a qualitative character also returned a significant relationship for PC1 (P < 0.002 across the three models). Parallel PGLS and ANOVA analyses with shape PC2 did not result in any significant effects of pollinators on this aspect of shape (Tables S2 and S3), consistent with the findings from the original dataset.

### PATTERNS OF CONVERGENCE

Given the apparent relationship between the aspects of shape captured by PC1 and pollination system, we examined the varia-



**Figure 2.** Biplots of hummingbird importance and PC axes. Each point represents one of the 16 species in the analysis; regression line from PGLS analysis. Hummingbird importance ranges from 0 (all insect pollination) to 1 (all hummingbird pollination); see Methods. The top panel shows results for PC1 and the bottom panel for PC2.

tion in PC1 across the phylogeny and tested for significant shifts in shape. Using the MCC tree, seven lineages are inferred to have shifted away from the ancestral regime (Fig. 3, Fig. S2), and many of these fall within the campanulate, bowl-shaped, and



**Figure 3.** Traitgram for floral shape PC1. Branches are colored by regimes estimated using 11ou on the MCC tree. Each color represents a unique regime, a distinct inferred selective optimum for shape PC1. Example floral shapes corresponding to the PC values are shown on the left.

funnel-shaped groups. Three lineages (Eriolarynx + Vassobia, S. punctata, and I. lehmannii) are estimated to have shifted independently to the same regime, and one clade (that includes D. spinosa) appears to have subsequently reverted to the ancestral tubular shape (Fig. 3, Fig. S2). Three lineages (I. grandiflorum, I. australe, and A. arborescens) are estimated to be evolving toward unique optima despite the fact that their values for PC1 are similar to taxa in other regimes. This result likely relates to the short length of the subtending branch relative to the observed trait value. For example, since I. australe has experienced such a large change in shape over a short period of time, the optimum toward that is it is evolving is inferred to be different than the other black-colored lineages with similar present-day values. Given this interaction between branch length variation and inferred shifts, we compared the number and placement of shifts across all of the 100 posterior trees. We found a slightly lower number of inferred shifts on average (4.83  $\pm$  1.22 SD compared to 7 on the MCC tree) and also fewer convergent shifts (2.28  $\pm$ 0.50 SD compared to 4 on the MCC tree) (Fig. 3, Fig. S2). Still, the overall patterns were similar across the analyses. For example, I. australe appeared as a shift in 94 of the 100 trees and this shift was consistently inferred to correspond to a distinct regime (Tables S3). Also, the reversion of the clade formed by D. spinosa, D. brachyacantha, and I. parvifolium to the ancestral regime was inferred in the 61% of the trees (Table S4).

# Discussion

The pollinator-shift model suggests that macroevolutionary transitions between groups of pollinators will lead to changes in multiple floral traits. Although this concept is long standing in the literature (Johnson 2006), empirical tests of this hypothesis, particularly with quantitative measures of pollination system, remain few (reviewed in Rausher 2008; Smith 2010). Some of the strongest evidence for a tight relationship between pollination system and floral trait evolution comes from intraspecific studies or studies of species pairs (e.g., Johnson and Steiner 1997; Perez-Barrales et al. 2007; Streisfeld and Kohn 2007). However, studies of entire clades have the potential to capture multiple independent transitions, providing greater power for hypothesis testing. Moreover, macroevolutionary analyses are crucial for addressing how often pollinator shifts are tied to speciation events (van der Niet and Johnson 2012).

Our results suggest that changes in corolla shape are often linked to changes in pollination system, consistent with pollinator shift model. In Iochrominae, species with tubular corollas tend to be pollinated primarily by hummingbirds, those with open corollas (bowl-shaped or campanulate) by insects, and those with funnel-shaped corollas by a mixture. The insect pollinators are principally bees except in *A. arborescens* and *I. ellipticum*, which have generalized systems with contributions from multiple hymenopterans, dipterans, and lepidopterans (Smith et al. 2008b). A wide variety of pollinators has been shown to exert selection on corolla shape in microevolutionary studies (e.g., Galen 1996; Gomez et al. 2006; Kulbaba and Worley 2013). Nonetheless, our study appears to be the first to examine this relationship on a macroevolutionary scale, treating both pollination system and floral shape as quantitative traits (see Gomez et al. (2015) for a discrete example).

The pattern of transitions inferred in Iochrominae reinforces the theme of reversibility in pollination ecology and associated traits (Vamosi et al. 2014). Iochrominae belongs to the Physalid clade of the tomato family, which is inferred to be ancestrally open-flowered and bee-pollinated (Knapp 2010). Tubular flowers have evolved many times in the family outside of the Physalids, and at least in some cases, these taxa are documented to be pollinated by hummingbirds, hawkmoths, or both (Sazima et al. 1995; Ippolito et al. 2004; Gubitz et al. 2009). Our reconstruction of shape evolution in Iochrominae demonstrates the potential for reevolution of open flowers with bee or generalist insect pollination in tubular clades (Fig. 3). This transition may be facilitated by the retention of at least some degree of insect pollination in most of the tubular taxa (Fig. 2). In Iochrominae, the exclusively insect pollinated taxa (V. breviflora, A. arborescens, I. ellipticum) are found at lower elevations (Smith 2006), and thus the reversal to open flowers is likely related to range expansions from the Andes into adjacent regions. Similar lability of floral traits and pollination systems has been documented in many clades (Perret et al. 2003; Tripp and Manos 2008; Marten-Rodriguez et al. 2010) although the relationship to range evolution is largely untested (Herrera et al. 2006).

While our study supports a close association between floral shape and pollination system, additional studies would be needed to determine the underlying drivers of this relationship. Pollinators could exert selection on shape either through their rates of visitation or through their effectiveness in pollen removal and deposition during those visits. Manipulative experiments in the principally bee-pollinated Penstemon strictus have shown that artificially constricting the corolla tube results in fewer bee visits (Zung et al. 2015), a shorter duration of these visits, and reduced pollen removal (Castellanos et al. 2004). The latter effects are likely related to aspects of mechanical fit, but the reduced visitation suggests that visual cues may also be important. Nectar guides are presumably more visible in flowers with a wider opening, and these patterns invoke innate preferences in bees (Lehrer et al. 1995). In this context, it is notable that the bowl-shaped and campanulate corollas in Iochrominae present dramatic patterns on the inside surface of the corolla while the tubular species are uniformly colored or at most, slightly patterned on the lobes. Experimental manipulations of these corolla patterns (as in Koski and Ashman 2014) as well as of the floral tube would help to

tease apart these trait interactions and determine how the aspects of shape variation captured in the geometric morphometric analysis alter interactions with pollinators in Iochrominae.

Together with previous work in this system, corolla form appears to be one of several floral traits, including nectar reward and floral display size, which fit the pollinator-shift model in Iochrominae. Although not quantitatively examined, floral scent may also belong to this suite of traits, as scent appears to be present only in the insect or mixed pollination taxa (Kaiser 2000; S. D. Smith, pers. obs.). Nonetheless, other traits, most notably flower color, do not correlate well with pollinator shifts (Smith et al. 2008a). Although the three red-flowered Iochroma species are principally pollinated by hummingbirds, the remaining hummingbird-pollinated taxa exhibit other colors (white, green, purple, yellow), and the geographical distribution of this color variation is consistent with a competition-driven model of floral divergence (Muchhala et al. 2014). These results support the notion that pollination syndromes are not likely to be universal (Ollerton et al. 2009; Ollerton et al. 2015), that is, that they are not likely to explain floral diversity across all traits or clades. In this context, the core question for future studies should not be whether the pollinator-shift model applies, as clearly it does in some cases, but rather when and why it is likely to apply. Addressing this question will require both microevolutionary studies to measure selective forces acting within lineages as well as macroevolutionary studies to detect broad patterns and overarching processes.

## AUTHOR CONTRIBUTIONS

S.D.S. and R.K. designed the research and conducted the analyses. S.D.S. wrote the manuscript with revisions by R.K.

#### ACKNOWLEDGMENTS

This work was supported by National Science Foundation grants to S.D.S. (NSF DEB-1413855 and 1355518). R.K. was supported by NSF DEB-1655611. The authors wish to thank C. Ané and M. Khabbazian for valuable advice on statistical analyses and two anonymous reviewers for constructive feedback.

#### DATA ARCHIVING

All input data (shape outlines, sequence alignments, phylogenetic trees) and scripts for analysis are available at Dryad repository (https://doi.org/10.5061/dryad.5jn7b).

#### LITERATURE CITED

- Armbruster, W. S. 1990. Estimating and testing the shapes of adaptive surfaces: the morphology and pollination of *Dalechampia* blossoms. Am. Nat. 135:14–31.
- 2002. Can indirect selection and genetic context contribute to trait diversification? A transition-probability study of blossom-colour evolution in two genera. J. Evol. Biol. 15:468–486.
- Armbruster, W. S., M. E. Edwards, and E. M. Debevec. 1994. Floral character displacement generates assemblage structure of Western-Australian triggerplants (*Stylidium*). Ecology 75:315–329.

- Barrett, S. C. H., and K. A. Hodgins. 2006. Floral design and the evolution of asymmetrical mating systems. Pp. 239–255 *in* L. D. Harder, and S. C. H. Barrett, eds. Ecology and evolution of flowers. Oxford Univ. Press, Oxford.
- Blomberg, S. P., T. Garland Jr., and A. R. Ives. 2003. Testing for phylogenetic signal in comparative data: behavioral traits are more labile. Evolution 57:717–745.
- Bonhomme, V., S. Picq, C. Gaucherel, and J. Claude. 2014. Momocs: outline analysis using R. J. Stat. Softw. 56:1–24.
- Bouckaert, R., J. Heled, D. Kuhnert, T. Vaughan, C. H. Wu, D. Xie, M. A. Suchard, A. Rambaut, and A. J. Drummond. 2014. BEAST 2: a software platform for Bayesian evolutionary analysis. Plos Comput. Biol. 10:e1003537.
- Butler, M. A., and A. A. King. 2004. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. Amer. Nat. 164:683–695.
- Campbell, D. R. 2004. Natural selection in *Ipomopsis* hybrid zones: implications for ecological speciation. New Phytol. 161:83–90.
- Campbell, D. R., N. M. Waser, and M. V. Price. 1996. Mechanisms of hummingbird-mediated selection for flower width in *Ipomopsis aggregata*. Ecology 77:1463–1472.
- Castellanos, M. C., P. Wilson, and J. D. Thomson. 2004. "Anti-bee" and "pro-bird" changes during the evolution of hummingbird pollination in *Penstemon* flowers. J. Evol. Biol. 17:876–885.
- Conner, J. K. 2006. Ecological genetics of floral evolution. Pp. 260–277 in L. D. Harder, and S. C. H. Barrett, eds. Ecology and evolution of flowers. Oxford Univ. Press, Oxford.
- Cresswell, J. E., and C. Galen. 1991. Frequency-dependent selection and adaptive surfaces for floral character combinations: the pollination of *Polemonium viscosum*. Am. Nat. 138:1342–1353.
- Davis, C. C., H. Schaefer, Z. Xi, D. A. Baum, M. J. Donoghue, and L. J. Harmon. 2014. Long-term morphological stasis maintained by a plantpollinator mutualism. Proc. Natl. Acad. Sci. USA 111:5914–5919.
- De Luca, P. A., and M. Vallejo-Marin. 2013. What's the "buzz" about? The ecology and evolutionary significance of buzz-pollination. Curr. Opin. Plant Biol. 16:429–435.
- Eaton, D. A. R., C. B. Fenster, J. Hereford, S. Q. Huang, and R. H. Ree. 2012. Floral diversity and community structure in *Pedicularis* (Orobanchaceae). Ecology 93:S182–S194.
- Fenster, C. B., W. S. Armbruster, P. Wilson, M. R. Dudash, and J. D. Thomson. 2004. Pollination syndromes and floral specialization. Annu. Rev. Ecol. Evol. S 35:375–403.
- Fenster, C. B., G. Cheely, M. R. Dudash, and R. T. Reynolds. 2006. Nectar reward and advertisement in hummingbird-pollinated *Silene virginica* (Caryophyllaceae). Am. J. Bot. 93:1800–1807.
- Fernandez-Hilario, R., and S. D. Smith. 2017. A new species of *Saracha* (Solanaceae) from the Central Andes of Peru. Phytokeys 85:31–43.
- Fernandez-Mazuecos, M., J. L. Blanco-Pastor, J. M. Gomez, and P. Vargas. 2013. Corolla morphology influences diversification rates in bifid toadflaxes (*Linaria* sect. Versicolores). Ann. Bot. 112:1705–1722.
- Galen, C. 1996. Rates of floral evolution: adaptation to bumblebee pollination in an alpine wildflower, *Polemonium viscosum*. Evolution 50:120–125.
- Galen, C., and J. Cuba. 2001. Down the tube: pollinators, predators, and the evolution of flower shape in the alpine skypilot, *Polemonium viscosum*. Evolution 55:1963–1971.
- Gomez, J. M., J. Bosch, F. Perfectti, J. D. Fernandez, M. Abdelaziz, and J. P. M. Camacho. 2008. Association between floral traits and rewards in *Erysimum mediohispanicum* (Brassicaceae). Ann. Bot. 101:1413–1420.
- Gomez, J. M., F. Perfectti, and J. P. M. Camacho. 2006. Natural selection on *Erysimum mediohispanicum* flower shape: insights into the evolution of zygomorphy. Am. Nat. 168:531–545.

- Gomez, J. M., F. Perfectti, and C. P. Klingenberg. 2014. The role of pollinator diversity in the evolution of corolla-shape integration in a pollinationgeneralist plant clade. Philos. T R Soc. B 369:20130257.
- Gomez, J. M., F. Perfectti, and J. Lorite. 2015. The role of pollinators in floral diversification in a clade of generalist flowers. Evolution 69:863–878.
- Grant, V. 1949. Pollination systems as isolating mechanisms in angiosperms. Evolution 3:82–97.
- Gubitz, T., M. E. Hoballah, A. Dell'Olivo, and C. Kuhlemeier. 2009. Petunia as a model system for the genetics and evolution of pollination syndromes.
  Pp. 29–50 *in* T. Gerats, and J. Strommer, eds. Petunia: Evolutionary, developmental and physiological genetics. Springer, New York.
- Hansen, T. F. 1997. Stabilizing selection and the comparative analysis of adaptation. Evolution 51:1341–1351.
- Herrera, C. M., M. C. Castellanos, and M. Medrano. 2006. Goegraphical context of floral evolution: towards an improved research programme in floral diversification. Pp. 278–294 *in* L. D. Harder, and S. C. H. Barrett, eds. Ecology and evolution of flowers. Oxford Univ. Press, Oxford.
- Ippolito, A., G. W. Fernandes, and T. P. Holtsford. 2004. Pollinator preferences for *Nicotiana alata*, *N. forgetiana*, and their F-1 hybrids. Evolution 58:2634–2644.
- Johnson, S. D. 2006. Pollinator-driven speciation in plants. Pp. 296–306 in L. D. Harder, and S. C. H. Barrett, eds. The ecology and evolution of flowers. Oxford Univ. Press, Oxford.
- Johnson, S. D., and K. E. Steiner. 1997. Long-tongued fly pollination and evolution of floral spur length in the *Disa draconis* complex (Orchidaceae). Evolution 51:45–53.

Kaiser, R. 2000. Scents from rain forests. Chimia 54:346–363.

- Khabbazian, M., R. Kriebel, K. Rohe, and C. Ané. 2016. Fast and accurate detection of evolutionary shifts in Ornstein-Uhlenbeck models. Methods Ecol. Evol. 7:811–824.
- Klingenberg, C. P. 2010. Evolution and development of shape: integrating quantitative approaches. Nat. Rev. Genet. 11:623–635.
- Knapp, S. 2010. On "various contrivances": pollination, phylogeny and flower form in the Solanaceae. Philos. T R. Soc. B 365:449–460.
- Koski, M. H., and T. L. Ashman. 2014. Dissecting pollinator responses to a ubiquitous ultraviolet floral pattern in the wild. Funct. Ecol. 28:868–877.
- Kulbaba, M. W., and A. C. Worley. 2013. Selection on *Polemonium brandegeei* (Polemoniaceae) flowers under hummingbird pollination: in ppposition, parallel, or independent of selection by hawkmoths? Evolution 67:2194– 2206.
- La Rosa, R. J., and J. K. Conner. 2017. Floral function: effects of traits on pollinators, male and female pollination success, and female fitness across three species of milkweeds (*Asclepias*). Am. J. Bot. 104:150–160.
- Lehrer, M., G. A. Horridge, S. W. Zhang, and R. Gadagkar. 1995. Shape vision in bees: innate preference for flower-like patterns. Philos. T R Soc. B 347:123–137.
- Marten-Rodriguez, S., C. B. Fenster, I. Agnarsson, L. E. Skog, and E. A. Zimmer. 2010. Evolutionary breakdown of pollination specialization in a Caribbean plant radiation. New Phytol. 188:403–417.
- Martins, E. P., and T. F. Hansen. 1997. Phylogenies and the comparative method: a general approach to incorporating phylogenetic information into the analysis of interspecific data. Am. Nat. 149:646–667.
- Mayfield, M. M., N. M. Waser, and M. V. Price. 2001. Exploring the "most effective pollinator principle" with complex flowers: bumblebees and *Ipomopsis aggregata*. Ann. Bot. 88:591–596.
- Muchhala, N. 2007. Adaptive trade-off in floral morphology mediates specialization for flowers pollinated by bats and hummingbirds. Am. Nat. 169:494–504.
- Muchhala, N., S. Johnsen, and S. D. Smith. 2014. Competition for hummingbird pollination shapes flower color variation in Andean Solanaceae. Evolution 68:2275–2286.

- Muchhala, N., and M. D. Potts. 2007. Character displacement among batpollinated flowers of the genus *Burmeistera*: analysis of mechanism, process and pattern. P R Soc. B 274:2731–2737.
- Ne'eman, G., A. Jurgens, L. Newstrom-Lloyd, S. G. Potts, and A. Dafni. 2010. A framework for comparing pollinator performance: effectiveness and efficiency. Biol. Rev. 85:435–451.
- Ollerton, J., R. Alarcon, N. M. Waser, M. V. Price, S. Watts, L. Cranmer, A. Hingston, C. I. Peter, and J. Rotenberry. 2009. A global test of the pollination syndrome hypothesis. Ann. Bot. 103:1471–1480.
- Ollerton, J., A. R. Rech, N. M. Waser, and D. Price. 2015. Using the literature to test pollination syndromes—some methodological cautions. J. Pollin. Ecol. 16:119–125.
- Perez-Barrales, R., J. Arroyo, and W. S. Armbruster. 2007. Differences in pollinator faunas may generate geographic differences in floral morphology and integration in *Narcissus papyraceus* (Alarcissiopapyraceris). Oikos 116:1904–1918.
- Perez, F., M. T. K. Arroyo, R. Medel, and M. A. Hershkovitz. 2006. Ancestral reconstruction of flower morphology and pollination systems in *Schizanthus* (Solanaceae). Am. J. Bot. 93:1029–1038.
- Perret, M., A. Chautems, R. Spichiger, G. Kite, and V. Savolainen. 2003. Systematics and evolution of tribe Sinningieae (Gesneriaceae): evidence from phylogenetic analyses of six plastid DNA regions and nuclear ncpGS. Am. J. Bot. 90:445–460.
- Pinheiro, J., D. Bates, S. DebRoy, D. Sarkar and R Core Team. 2017. nlme: Linear and Nonlinear Mixed Effects Models. R package version 3:1– 131, https://CRAN.R-project.org/package=nlme.
- Rausher, M. D. 2008. Evolutionary transitions in floral color. Int. J. Plant Sci. 169:7–21.
- Revell, L. J. 2012. Phytools: an R package for phylogenetic comparative biology (and other things). Methods Ecol. Evol. 3:217–223.
- Revell, L. J., L. J. Harmon, and D. C. Collar. 2008. Phylogenetic signal, evolutionary process, and rate. Syst. Biol. 57:591–601.
- Rosas-Guerrero, V., R. Aguilar, S. Marten-Rodriguez, L. Ashworth, M. Lopezaraiza-Mikel, J. M. Bastida, and M. Quesada. 2014. A quantitative review of pollination syndromes: do floral traits predict effective pollinators? Ecol. Lett. 17:388–400.
- Särkinen, T., L. Bohs, R. G. Olmstead, and S. Knapp. 2013. A phylogenetic framework for evolutionary study of the nightshades (Solanaceae): a dated 1000-tip tree. BMC Evol. Biol. 13:214.
- Sazima, I., S. Buzato, and M. Sazima. 1995. An assemblage of hummingbirdpollinated flowers in a montane forest in southeastern Brazil. Bot. Acta 109:149–160.
- Smith, S. D. 2006. Floral diversification and pollination biology of the Andean clade Iochrominae (Solanaceae). Department of Botany, University of Wisconsin, Madison.
- Smith, S. D., C. Ane, and D. A. Baum. 2008a. The role of pollinator shifts in the floral diversification of *Iochroma* (Solanaceae). Evolution 62:793– 806.
- Smith, S. D., and D. A. Baum. 2006. Phylogenetics of the florally diverse Andean clade Iochrominae (Solanaceae). Am. J. Bot. 93:1140–1153.
- 2007. Systematics of Iochrominae (Solanaceae): patterns in floral diversity and interspecific crossability. Acta Horticulturae 745:241–254.
- Smith, S. D., and E. E. Goldberg. 2015. Tempo and mode of flower color evolution. Am. J. Bot. 102:1014.
- Smith, S. D., S. J. Hall, P. R. Izquierdo, and D. A. Baum. 2008b. Comparative pollination biology of sympatric and allopatric Andean *Iochroma* (Solanaceae). Annal. Missouri Bot. Garden 95:600–617.

- Stebbins, G. L. 1970. Adaptive radiation of reproductive characteristics in angiosperms, I: pollination mechanisms. Annu. Rev. Ecol. Syst. 1:307– 326.
- Strauss, S. Y., and J. B. Whittall. 2006. Non-pollinator agents of selection on floral traits. Pp. 120–138 in S. C. H. Barrett, and L. D. Harder, eds. The ecology and evolution of flowers. Oxford Univ. Press, Oxford.
- Streisfeld, M. A., and J. R. Kohn. 2007. Environment and pollinator-mediated selection on parapatric floral races of *Mimulus aurantiacus*. J. Evol. Biol. 20:122–132.
- Sutherland, S. D., and R. K. Vickery. 1996. On the relative importance of flower color, shape, and nectar rewards in attracting pollinators to *Mimulus*. Great Basin Nat. 56:282–282.
- Swofford, D. L. 2002. PAUP\*: phylogenetic analysis using parsimony (\*and other methods). Sinauer, Sunderland, Massachusetts.
- Taura, H. M., and S. Laroca. 2004. Biologia da polinização: interações entre as abelhas (Hym., Apoidea) e as flores de Vassobia breviflora (Solanaceae). Acta Biológica Paranaense 33:143–162.
- Temeles, E. J., C. R. Koulouris, S. E. Sander, and W. J. Kress. 2009. Effect of flower shape and size on foraging performance and trade-offs in a tropical hummingbird. Ecology 90:1147–1161.
- Tibshirani, R. 1996. Regression shrinkage and selection via the lasso. J. R. Stat. Soc. Ser. B Methodology 58:267–288.
- Tripp, E. A., and P. S. Manos. 2008. Is floral specialization an evolutionary dead-end? Pollination system transitions in *Ruellia* (Acanthaceae). Evolution 62:1712–1736.
- Vamosi, J. C., W. S. Armbruster, and S. S. Renner. 2014. Evolutionary ecology of specialization: insights from phylogenetic analysis. P R Soc. B 281:20142004.
- van der Niet, T., and S. D. Johnson. 2012. Phylogenetic evidence for pollinatordriven diversification of angiosperms. Trends Ecol. Evol. 27:353–361.
- Van der Niet, T., R. Peakall, and S. D. Johnson. 2014. Pollinator-driven ecological speciation in plants: new evidence and future perspectives. Ann. Bot. 113:199–211.
- van der Niet, T., C. P. E. Zollikofer, M. S. P. de Leon, S. D. Johnson, and H. P. Linder. 2010. Three-dimensional geometric morphometrics for studying floral shape variation. Trends Plant Sci. 15:423–426.
- Verçoza, F., A. R. Dias, and C. C. Missagia. 2012. Ecologia da polinização e potenciais dispersores da "marianeira"—*Acnistus arborescens* (L.) Schltdl. (Solanaceae) em área de Floresta Atlântica do Rio de Janeiro. Natureza On Line 10:59–64.
- Waser, N. M., L. Chittka, M. V. Price, N. M. Williams, and J. Ollerton. 1996. Generalization in pollination systems, and why it matters. Ecology 77:1043–1060.
- Waser, N. M., and M. V. Price. 1983. Optimal and actual outcrossing in plants, and the nature of plant-pollinator interaction. Pp. 341–359 *in* C. E. Jones, and R. J. Little, eds. Handbook of experimental pollination biology. Van Nostrand Reinhold, New York.
- Whittall, J. B., and S. A. Hodges. 2007. Pollinator shifts drive increasingly long nectar spurs in columbine flowers. Nature 447:706–709.
- Wilson, P., M. C. Castellanos, J. N. Hogue, J. D. Thomson, and W. S. Armbruster. 2004. A multivariate search for pollination syndromes among penstemons. Oikos 104:345–361.
- Zung, J. L., J. R. K. Forrest, M. C. Castellanos, and J. D. Thomson. 2015. Beeto bird-pollination shifts in Penstemon: effects of floral-lip removal and corolla constriction on the preferences of free-foraging bumble bees. Evol. Ecol. 29:341–354.

## Associate Editor: M. Vallejo-Marin Handling Editor: M. Servedio

# Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's website:

Figure S1. Timetree of Iochrominae. Maximum clade credibility tree from BEAST analysis.

95% highest posterior densities for each node depth are shown with the light blue bars.

Branches with less than 95% posterior probability (PP) are colored in gray and those with greater PP values are in black. Time is indicated in millions of years (mya) along the x-axis.

**Figure S2.** Estimate of regime shifts on MCC tree. Significant shifts are indicated with black asterisks and can be either transitions to new shape regimes or reversals to the ancestral regime. Distinct regimes are indicated with different colors; here there are 5 estimated regimes. Analysis is based on PC1 values, plotted on the right.

**Table S1.** Pollinator data for Iochrominae taxa. Quantitative measures of relative hummingbird importance are listed where available. For the remaining taxa, principal pollinator was coded from field observations. Voucher specimens are listed for personal observations.

**Table S2.** Parameter estimates from PGLS analyses. "Poll" denotes relative hummingbird importance and the PCs correspond to the shape PCs (Fig. 1). The "No Vb" analysis has the outlier *Vassobia breviflora* removed from the dataset (see text).

Table S3. Phylogenetic ANOVA with pollinator system as a categorical variable. "Poll" denotes relative hummingbird importance and the PCs correspond to the shape PCs (Fig. 1).

**Table S4.** Inclusion in a regime shift. Each species is listed along with the frequency with which it was associated with a shift across the 100-tree sample. **Table S5.** Distinct convergent regimes estimated by 11ou across the 100-tree sample. These include both cases where independent lineages acquired the same derived regime (e.g., the blue lineages in Fig. S2) and where lineages reverted to the ancestral (background) regime (e.g., the *D. spinosa*+ *D. brachyachantha*+*I. parvifolium* clade in Fig. S2).